YALE ANIMAL RESOURCES CENTER BSL-2 PROCEDURES

The YARC will follow established regulatory policies. The outline listed below is the standard policy for handling of animals infected with BSL-2 agents. Biological safety may advise additional procedures on a case by case basis, for larger animals or unique circumstances.

NOTE: PI must contact the YARC Facility Supervisor (72 hours-3 business days) <u>prior</u> to administering compound to ensure YARC husbandry staff are advised of the use of a (potentially) hazardous agent.

Labeling

- Animal room door must be labeled with a biohazard symbol and a BSL-2 sign with all information completed on the sign.
- ☐ Cages must be labeled with the "BSL-2 Agent in Use" cage card. All information must be completed by the P.I./researcher (agent, PI, day and emergency contact phone #s)

Protective clothing

- Solid front, long sleeve gown with gloves (recommend double glove) pulled over the gown sleeve cuff.
 - Contaminated gown is removed in animal room and deposited in appropriate container.
- ☐ Gloves should be removed, hands washed thoroughly before exiting room.
- A mask and eye protection must be worn if a Class II biosafety cabinet (BSC) is not available.
- ☐ Biological Safety will advise if additional personal protection is required (i.e. respirator).

Engineering Controls

- A BSC is required for the handling of (small) animals and experimental manipulations. Biological safety will advise if not available or for unique circumstances.
 - Air pressure differential of the animal room must be set at negative pressure.

Caging

Static Micro-isolator (MI) with filter top. Biological safety will advise if alternative is acceptable.

Work Space/Practices

- ☑ Once animals are infected, cages should only be opened in the BSC.
- Decontaminate the BSC with MB-10 or approved disinfectant before and after use; allow 3-10 minutes contact time.
- Assess area prior to set up; ensure sharps containers are in the BSC
- Place cage on disinfectant soaked paper towel prior to opening top and working with animals.
- Disinfect outside of cages prior to removal from the BSC by wiping with MB-10 or approved disinfectant soaked paper towel avoiding filter top (do not spray)
- ☑ Other work surfaces can be decontaminated with 10% bleach, MB-10, or approved disinfectant
- Husbandry & experimental procedures should be conducted in such manner to minimize the potential release of aerosols into the environment (i.e. do not vacuum any BSL-2 waste or
- materials, caution when handling to prevent spills)
 - ☑ Room sanitizing agent is Saniplex.

Experimental Manipulations

- ☑ Use a separate syringe and needle for each animal.
- Dispose of sharps immediately in sharps container; do not rest down on work surface.
- ☐ Reusable sharps and instruments should be placed in a solid container for personnel safety.
- Soak in 10% bleach solution for 30 minutes and rinse well with ethanol.

Decontamination

Caging

- ☐ The MI filter bonnet must remain on the soiled cage to minimize potential for contamination.
- ☐ The BSL-2 cage card must also remain on the cage after animals are removed and cage is placed on the soiled cage rack.
- In most instances, there will be a designated cart or rack in the animal room for soiled cages.

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- Cages cannot be power-sprayed prior to placement in rack washer or tunnel washer.
- ☐ Caging must be washed in rack washer or tunnel washer at 180°f cycle for a minimum of 3 minutes.

<u>Waste</u>

- ☐ Carcasses and soft waste are bagged in biohazard bags within the BSC.
- ✓ Wrap carcasses in opaque absorbent material and label with PI and agent.
- ☐ Disinfect the outside of the bag prior to removal from the BSC.
- ☐ Change contaminated gloves prior to handling outside of the bag.
- ☐ Place in the BSL-2 bin in the facility cooler. Material may not be stored anywhere else.

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- All waste is autoclaved for 30 minutes at 250°f at 15 psi.
- After autoclaving, bedding is disposed of by YARC. Carcasses and soft waste are packaged and sent out as regulated medical waste.

<u>Transportation of Supplies and Animals</u>

- ☑ Place supplies in a primary leak proof container, then in secondary containment lined with absorbent material.
- ☐ Tubes and containers must be labeled with biohazard sticker with PI, agent, and phone number.
- ☐ In the event of a spill, the spill should be contained, the area marked off and either YARC (#785-2526) (if within animal facility) and/or Biosafety Office notified (#785-3555)
- ☐ Recommend carrying extra absorbent material during transport to contain a spill.
- ☐ Cages are to be secured with cargo straps and placed in secondary container

Emergency Response

- ✓ Secure animal(s)
- Apply first aid: Forced bleed and wash or flush mucous membranes.
- Review the *Procedures to Follow for Injuries Associated with Laboratory Animals:*https://your.yale.edu/policies-procedures/other/4438-policy-and-procedure-follow-injuries-associated-laboratory-animals
- ☑ Contact Employee Health #432-7978 or #432-0071 and EHS #785-3555 or 911 after hrs.

Medical Surveillance

- Case by case assessment of the agent and personal health history must be conducted by the Employee Health Department and Biosafety. Sera storage, consultations, sampling or immunizations may be recommended.
- Immunosuppressed or immunocompromised individuals should not be assigned to this project without consultation with the Employee Health Physician. Any individual with an immunosuppressive condition must notify the Employee Health Office (#432-7978 or #432-0071) for a confidential evaluation to request consideration for clearance to work on this project.
- If the health status of any individual assigned to this project changes after the initiation of the project, they must also notify the Yale Employee Health Office for a medical clearance review.

Please view Animal Biosafety Level 2 video at http://ehs.yale.edu/animal-biosafety

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